

Whole-Genome Sequence of *Enterobacter* sp. Strain SST3, an Endophyte Isolated from Jamaican Sugarcane (*Saccharum* sp.) Stalk Tissue

Han Ming Gan, a Sean E. McGroty, Teong Han Chew, Kok Gan Chan, Larry J. Buckley, Michael A. Savka, and André O. Hudson

ScienceVision SB, Setia Alam, Shah Alam, Selangor, Malaysia^a; The Thomas H. Gosnell School of Life Sciences, Rochester Institute of Technology, Rochester, New York, USA^b; Department of Biological Sciences, Faculty of Bioscience and Bioengineering, Universiti Teknologi Malaysia, Skudai, Johor, Malaysia^c; and Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, Kuala Lumpur, Malaysia^d

Enterobacter sp. strain SST3 is an endophytic bacterium isolated from Saccharum spp. Here we present its annotated draft genome that may shed light on its role as a bacterial endophyte of sugarcane. To our knowledge, this is the first genome announcement of a sugarcane-associated bacterium from the genus Enterobacter.

Sugarcane is an important crop plant that contributes to the economy of many countries, including Jamaica. Due to its economic importance, research addressing the growth and protection of sugarcane is a priority. There are examples in the literature of sugarcane research from countries such as Brazil and India that pertain to bacteria (8, 9). However, the same is not true for Jamaica even though the sugarcane industry is integral to Jamaica's economy.

Besides sugar production, sugarcane is valuable for other commodities, such as molasses, press mud (a solid residue before sugar crystallization), and biogases (fibrous matter that remains after juice extraction). In addition, this crop has recently garnered significant attention as perhaps the most economically competitive source for ethanol production (1).

The sequenced bacterium (SST3, for sugarcane stalk tissue) was isolated from an endophytic screen of stem tissues of sugarcane obtained from a farm in Savanna La Mar in the Westmoreland parish of Jamaica. SST3 was initially identified as an Enterobacter sp. strain by sequence analysis of the variable 3 region of the 16S rRNA gene (3). Strain SST3 was shown to produce bacterial cell-to-cell communication signals of the acyl-homoserine-lactone (AHL) family (3).

Nitrogen fertilization for sugarcane growth is very costly. As such, we are interested in the identification of sugarcane-associated nitrogen-fixing bacteria. A recent study showed that two *Enterobacter* strains from sugarcane roots and rhizosphere soil from China increase the biomass and nitrogen content of sugarcane seedlings to facilitate growth and development (6). The identification, sequencing, and annotation of the genome of this Jamaican sugarcane-associated bacterium have the potential to broaden our understanding of the role(s) that endophytic bacteria have in sugarcane growth and development.

The genome sequencing of *Enterobacter* sp. SST3 was performed using the Illumina MiSeq (150-bp paired-end reads). The reads were trimmed and assembled *de novo* using CLC Genomics Workbench 5.0 (CLC Bio, Denmark). Prodigal 2.60, tRNAscan-SE 1.3, and RNAmmer 1.2 were used to predict open reading frames (ORFs), tRNAs, and rRNAs, respectively (4, 5, 7). Subsequent genome annotation was performed using Blast2GO 2.5.1 (2). The *de novo* assembly results in 54-fold coverage of a 4,626,853-bp draft genome contained in 63 contigs with an average GC content of 56.05%. The contig N_{50} was approximately 150 kb, and the

longest contig assembled was approximately 330 kb. A total of 4,267 ORFs, 73 tRNAs, and 3 rRNAs were predicted from the draft genome.

Consistent with the AHL-positive phenotype of strain SST3, an AHL synthase gene was identified at contig 40. The protein sequence of this AHL synthase displays 88% similarity to CroI from *Citrobacter rodentium* strain CC168. Strain SST3 contains the complete enzyme set required for the conversion of sucrose into an energy source. Interestingly, an *iaaH* ortholog which may be involved in the production of auxin-like compounds that contribute to the promotion of sugarcane growth and development was also identified in the genome of strain SST3.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number ALNS000000000. The version described in this paper is the first version, ALNS01000000.

ACKNOWLEDGMENTS

This work was supported by a United States National Science Foundation (NSF) award to A.O.H. (MCB-1120541).

A.O.H., L.J.B., and M.A.S. thank the Thomas H. Gosnell School of Life Sciences and The College of Science at the Rochester Institute of Technology for ongoing support. We thank Carlton Scott for graciously providing sugarcane samples to facilitate this study.

REFERENCES

- 1. Buckeridge MS, De Souza AP, Arundale RA, Anderson-Teixeira KJ. 2012. Ethanol from sugarcane in Brazil: a "midway" strategy for increasing ethanol production while maximizing environmental benefits. Glob. Change Biol. Bioenergy 4:119–126.
- Conesa A, et al. 2005. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. Bioinformatics 21:3674

 3676
- 3. Hudson AO, Ahmad NH, Van Buren R, Savka MA. 2010. Sugarcane and grapevine endophytic bacteria: isolation, detection of quorum sensing signals and identification by 16S v3 rDNA sequence analysis, p 801–806. *In* Mendez-Vilas A (ed), Current research, technology and education topics in

Received 14 August 2012 Accepted 17 August 2012 Address correspondence to André O. Hudson, aohsbi@rit.edu. Copyright © 2012, American Society for Microbiology. All Rights Reserved. doi:10.1128/JB.01469-12

- applied microbiology and microbial biotechnology, vol 2. Formatex Research Center, Badajoz, Spain.
- 4. Hyatt D, et al. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119.
- Lagesen K, et al. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100 –3108.
- Lin L, et al. 2012. Plant growth-promoting nitrogen-fixing Enterobacteria are in association with sugarcane plants growing in Guangxi, China. Microbes Environ. [Epub ahead of Print]. http://dx.doi.org/10.1264/jsme2.ME11275.
- Lowe TM, Eddy SR. 1997. tRNA-scan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964.
- 8. Magnani GS, et al. 2010. Diversity of endophytic bacteria in Brazilian sugarcane. Genet. Mol. Res. 9:250–258.
- 9. Viswanathan R, Rajitha R, Sundar AR, Ramamoorthy V. 2003. Isolation and identification of endophytic bacteria strains from sugarcane stalks and their *in vitro* antagonism against the red rot pathogen. Sugar Tech. 5:25–29.

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